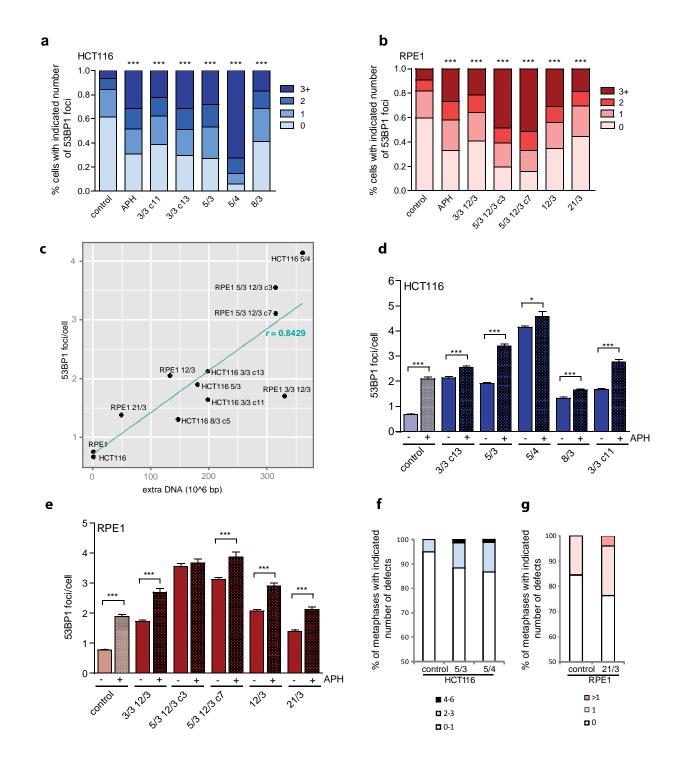


Supplementary Figure 1 Chromosome missegregation in trisomic and tetrasomic cells

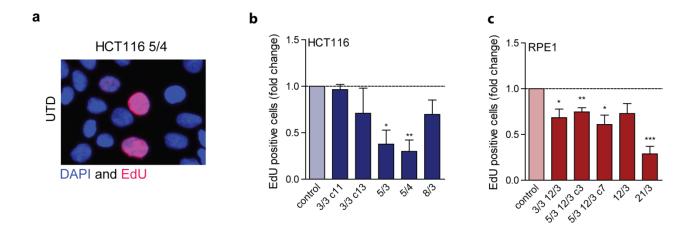
(a) Example of anaphase cell with a lagging chromosome. Bar = 10 μ m (b) (c) Quantification of lagging chromosomes in diploid controls and the respective trisomic and tetrasomic derivatives. Plots show mean \pm SEM of three independent experiments. At least 100 anaphases were scored in each experiment. (d) Example of anaphase cell with a lagging chromosome. Bar = 5 μ m. (e) Percentage of cells with lagging chromosomes scored as DNA mass positive for CREST staining. Plots show mean + SEM of three independent experiments. Non-parametric T-test.



Supplementary Figure 2 Trisomy and tetrasomy elevates DNA damage

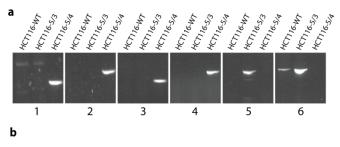
(a) (b) Quantification of % of cells with specific numbers of 53BP1 foci in HCT116 and the trisomic and tetrasomic derivatives (a) and RPE1 and the trisomic and tetrasomic derivatives (b). Control - parental cell line. APH -parental cell line treated with aphidicolin. Contingency tables were created from 3 independent experiments (n>500) and chi-square test was calculated comparing the number of cells with less than 3 foci or 3 and more foci in control and each trisomic and tetrasomic derivative. (c) The number of 53BP1 foci per cell scales with the amount of additional DNA. Note the similarity of the

independent clonal cells lines with the same extra chromosome - HCT116 3/3 c11 and c13 and RPE1 5/3 12/3 c3 and c7. (\mathbf{d})(\mathbf{e}) 53BP1 foci in parental and derived cell lines upon treatment with aphidicolin. Plots show mean \pm SEM of three independent experiments, at least 1000 cyclin A-negative cells were scored in each experiment. Non-parametric T test; ***: P<0.001. (\mathbf{f}) (\mathbf{g}) Occurence of metaphases with chromosome breaks and other abnormalities in untreated cells. N = 80,78,90,110,76 metaphases for HCT116, 5/3, 5/4, RPE1 and 21/3, respectively, obtained in two independent experiments.



Supplementary Figure 3 Sensitivity of aneuploid cells to replication stress

(a) Representative images of HCT116 5/4 cells stained with DAPI and EdU. Cells were grown in the presence of EdU for two hours. (**b**)(**c**) Quantification of EdU-positive cells in control HCT116 and aneuploid derivatives (**b**) and control RPE1 cells and aneuploid derivatives (**c**). All plots show mean \pm SEM of three independent experiments, at least 1000 cells were scored in each experiment. Non-parametric T test; * p<=0.05, ** p<=0.01, *** p<=0.001



No1: tandem duplication chr1:33540798-32841212

No2: deletion chr5:12286380-12291255

No3: tandem duplication chr7:30971351-30688299

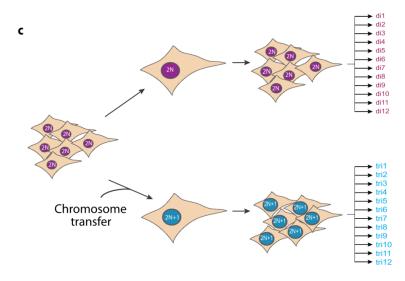
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No4: tandem duplication chr22:43949662-43930557

Junction:TTGGAAGGCCATGAAGGGCCATAGCGGCCCAGGGCTGCAGGTGGGCtACATACCATCTTGTGTGCATTCTGGATCTTCA
Chr22: TTGGAAGGCCATGAAGGGCCATAGCGGCCCAGGGGAGCAGGTTGTCGGAGCCTTTGAGCCACAGAAGAGCTCAGTCCAG
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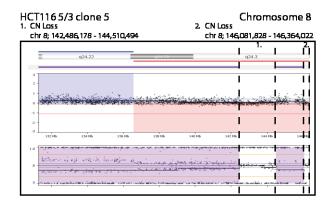
No5: inversion chr5: 88975284-88976028

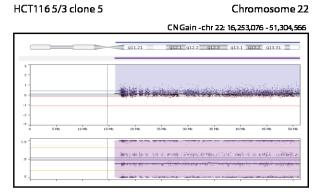
Junction: ATGACATATTAAAAGTGCAGAAGAAAAAAAAAATTACCCTAGTTTTTTCATCCATTCAGCAAGTCTATGTCTTGATGAGAGAGTTTAT
Chr5: ATGACATATTAAAAGTGCAGAAGAAAAAAAAAAATTACCCTAGAATAGTATATCTGGCAAAAATATCCTTCAAACATAAAGG
Chr5: ATAGGTGAAGTGTGTTTCTTGTAGACAACAGATGAATGGGTCTTGTTTTTTTCATCCATTCAGCAAGTCTATGTCTTGATTGGAGAGAGTTTAT

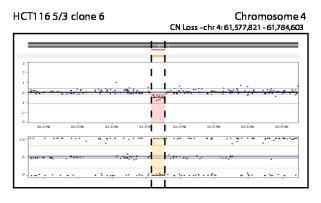


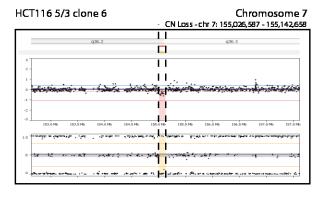
Supplementary figure 4 Mate-pair sequencing and single nucleotide polymorphism arrays to identify chromosomal rearrangements in an euploid cell lines

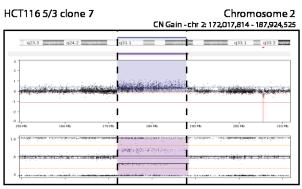
(a) Analysis of breakpoint junction by PCR for each of the six predicted de novo chromosomal rearrangements. Note that rearrangement 6 was also identified in the parental HCT116 indicating that this rearrangement did not occur de novo. (b) Breakpoint junction sequences of five de novo CNAs identified in aneuploid cells. In each case, the upper sequence denotes the breakpoint junction sequence and the two lower sequences denote the genomic regions where both flanks (red and blue) map to. Microhomologies are indicated in green. (c) Schematic depiction of the generation of clones derived from single cells that were used for the single nucleotide polymorphism (SNO) profiling. di1 - 12 clones derived from parental cell line HCT116; tri1 -12 clones derived from trisomic cell line HCT116 5/3. Two sets of these experiments were performed, 2x12 clonal cell populations were analyzed for each cell line.

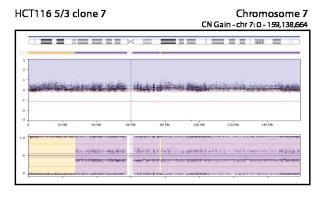


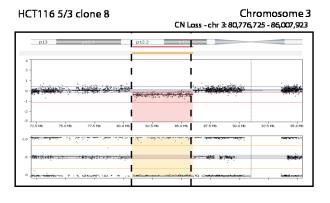


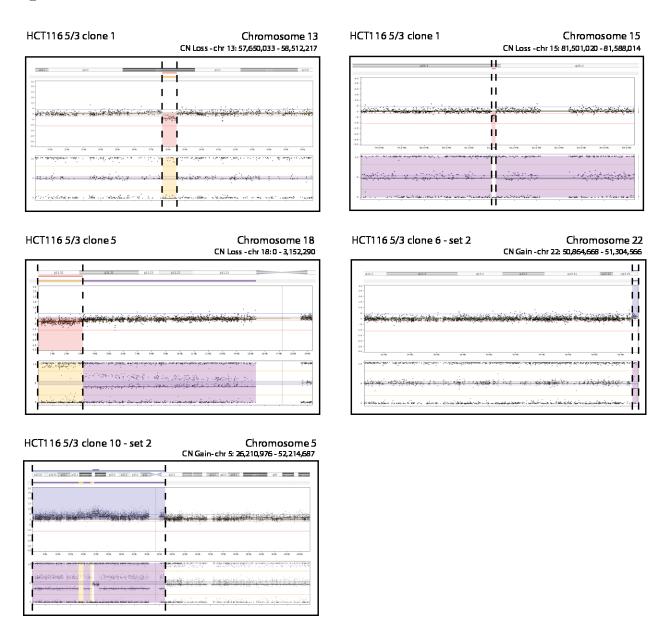






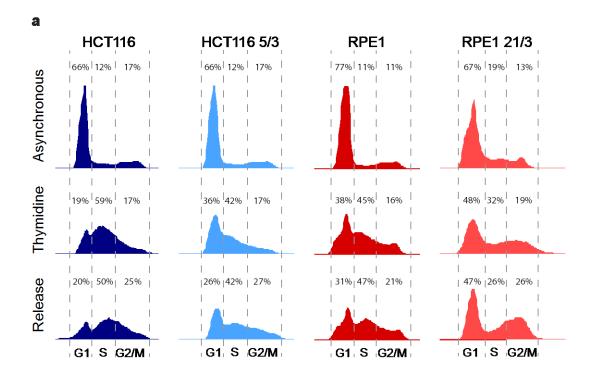






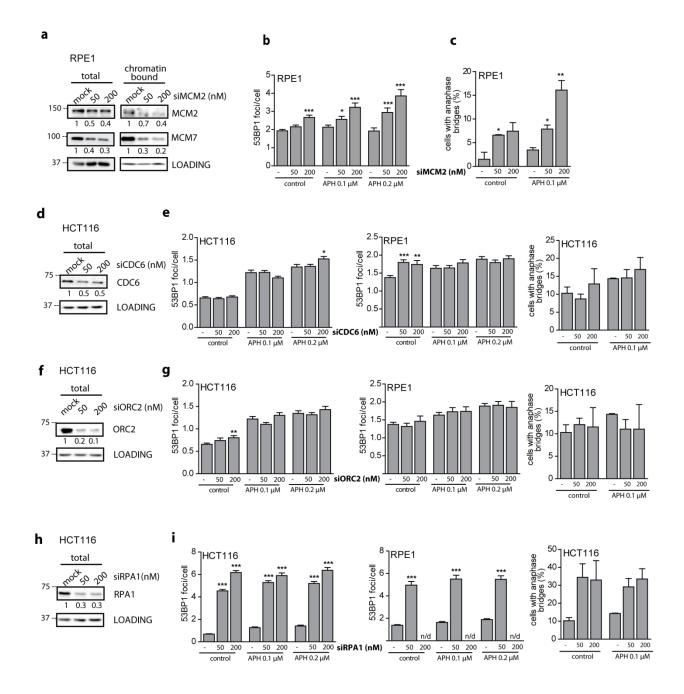
Supplementary Figure 5 Copy number alterations in trisomic cells

Unique *de novo* copy number alterations determinated in the individual clones derived from single aneuploid cell lines. Two independent sets of 12 single-cell clones were analyzed. (**a**) shows the CNAs from set 1; (**b**) set 2. Blue - copy number gain, red - copy number loss. Yellow and magenta denote the alleles A and B, respectively.



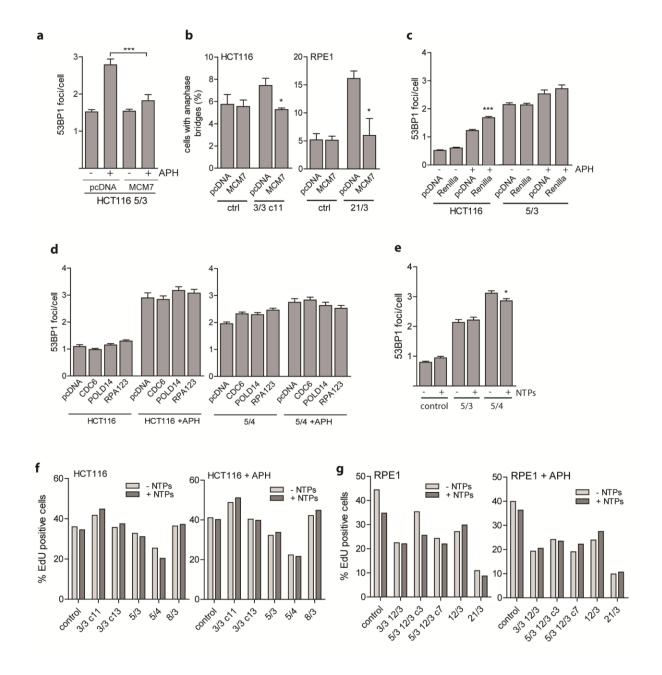
Supplementary Figure 6 Cell cycle profiles after thymidine synchronization and release

(a) Cell cycle profile of HCT116, HCT116 5/3, RPE1 and RPE1 21/3 under normal conditions (asynchronous), 30 hours after thymidine addition (thymidine) and 2 hours after thymidine washout (release).



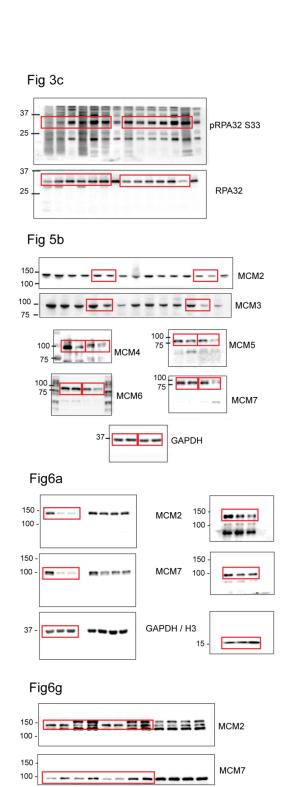
Supplementary Figure 7 Levels of replication proteins in aneuploid cells and the effects of downregulation of replication proteins in control cells

(a) Immunoblotting of whole cell lysates and chromatin bound fractions upon partial depletion of MCM2 by siRNA in the parental RPE1 cell line. Note the coordinate decrease in MCM7 abundance. (b) Average number of 53BP1 foci and (c) average number of anaphase bridges in cells depleted for MCM2. Partial depletion of CDC6 (d), ORC2 (f) and RPA1 (h) by siRNA in the parental HCT116 and RPE1 cell lines. Average number of 53BP1 foci and % of cells with anaphase bridges in cells depleted for CDC6 (e), ORC2 (g) and RPA1 (i). At least two independent experiments were performed and at least 500 cyclin A2-negative or 50 anaphases were scored for 53BP1 foci or anaphase bridge quantification, respectively. All plots show mean ± SEM; non-parametric T-test; * p<=0.05, ** p<=0.01, *** p<=0.001.



Supplementary Figure 8 The effects of NTP supplementation on EdU incorporation and the occurence of 53BP1 foci in aneuploid cells

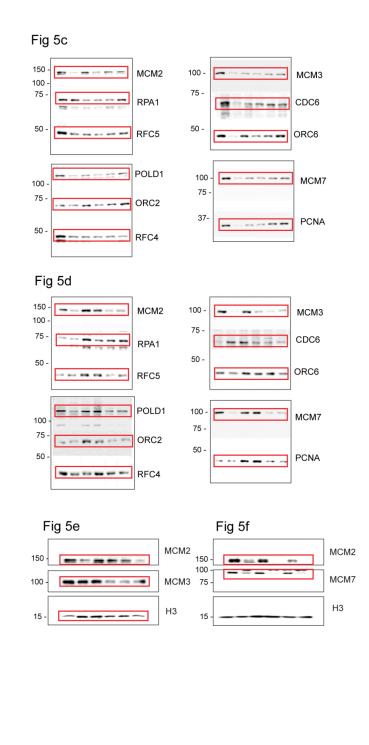
- (a) Accumulation of 53BP1 foci and (b) anaphase bridges upon transient overexpression of MCM7. One representative plot of three independent experiments (a) or mean ± SEM of three independent experiments (b) is shown. Non-parametric two-sided T test; *: P<0.05, **: P<0.01, ***: P<0.001.
- (c) Accumulation of 53BP1 foci in HCT116 and in HCT116 5/3 upon overexpression of Renilla luciferase. (d) Accumulation of 53BP1 foci in HCT116 and in HCT116 5/4 upon overexpression of replication factors. (e) 53BP1 foci formation in HCT116 and its aneuploid derivatives in the presence or absence of nucleoside supplement. Plot shows the average number of 53BP1 foci of at least 500 cyclin A2-negative cells collected in one experiment. Non-parametric t-test. * p<=0.05% EdU positivity in control HCT116 and aneuploid derivatives (f) and RPE1 and aneuploid derivatives (g) with or without nucleoside supplement (NTPs).



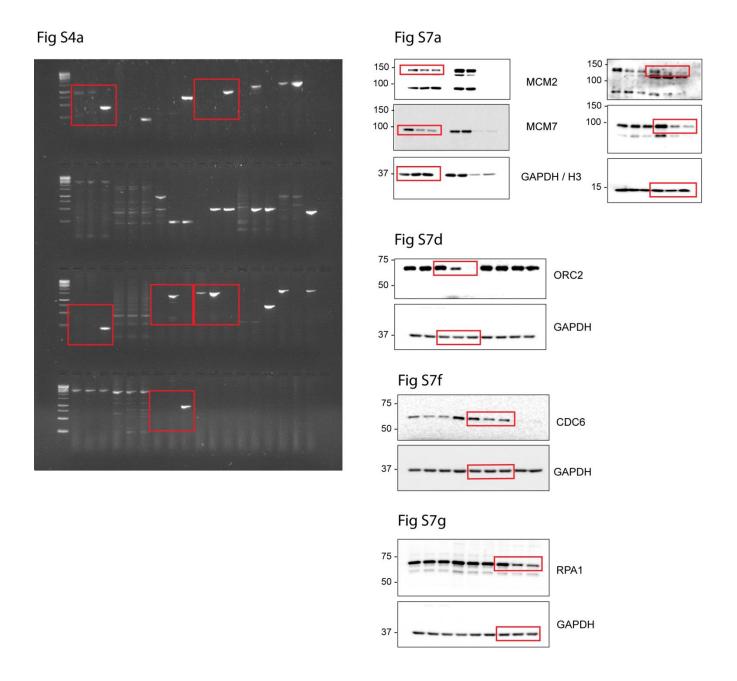
GAPDH

НЗ

15 -



Supplementary Figure 9 Uncropped blots of main figures Frames specify shown bands in indicated main figures.



Supplementary Figure 10 Uncropped blots of supplementary figures Frames specify shown bands in indicated supplementary figures.

Supplementary Table 1 List of all cell lines used in the analysis. % of cells with trisomy/tetrasomy was determined by chromosome painting. Note that the cell lines from Koi laboratory were used only for the analysis of the global proteome changes (Figure 5a).

Cell line name used in the text	Origin	Full cell line name	Analysis	% of trisomy or tetrasomy	Remarks
HCT116	HCT116 from AATC introduction H2B-GFP	HCT116 H2B-GFP	SNParrays CGH	-	Kuffer et al, 2013
HCT116 3/3 c11	MMTC into HCT116 H2B-GFP	HCT116 H2B-GFP 3/3 clone 11	SNParrays CGH	92 %	This work
HCT116 3/3 c13	MMTC into HCT116 H2B-GFP	HCT116 H2B-GFP 3/3 clone 13	SNParrays CGH	85 %	This work
HCT116 5/3	MMTC into HCT116 H2B-GFP	HCT116 H2B-GFP 5/3 clone 6	SNParrays CGH	92 %	Stingele et al, 2012
HCT116 5/4	MMTC into HCT116 H2B-GFP	HCT116 H2B-GFP 5/4 clone 4	SNParrays CGH	83 %	Stingele et al, 2012
HCT116 8/3	MMTC into HCT116 H2B-GFP	HCT116 H2B-GFP 8/3 clone 1	SNParrays CGH	78 %	Donnelly et al, 2014
RPE1	Taylor laboratory	RPE1 hTERT	SNParrays CGH	-	Kindly provided by Steven Taylor
RPE1 3/3 12/3	MMTC into RPE1		SNParrays CGH	100 %	This work Spontaneous gain of chromosome 12
RPE1 5/3 12/3 c3	MMTC into RPE1	RPE1 5/3 12/3 clone 3	SNParrays CGH	95 %	Stingele et al, 2012 Spontaneous gain of chromosome 12
RPE1 5/3 12/3 c7	MMTC into RPE1	RPE1 5/3 12/3 clone 7	SNParrays CGH	95 %	This work Spontaneous gain of chromosome 12
RPE1 12/3	Spontaneously arising trisomy of chromosome 12		CGH	100 %	This work Spontaneous gain of chromosome 12
RPE1	Taylor laboratory	RPE1 H2B-GFP hTERT	SNParrays CGH	-	Kindly provided by Steven Taylor
RPE1 21/3	MMTC into RPE1 H2B-GFP	RPE1 H2B-GFP 21/3	SNParrays CGH	90 %	Stingele et al, 2012
HCT116	Koi laboratory		SNParrays CGH	-	Kindly provided by Minoru Koi Haugen et al, 2008
HCT116 5/4	Koi laboratory		SNParrays CGH	88 %	Kindly provided by Minoru Koi Haugen et al, 2008
HCT116 3/3	Koi laboratory		CGH	82 %	Kindly provided by Minoru Koi Haugen et al, 2008

Supplementary Table 2 Overview of the identified copy number aberrations in HCT116 5/3.

Sample	chr	start	end	event	Size (bp)	Chr. band	Mosaic	Fragile site overlap
tr_13_set_2_clone_1	13	57650033	58512217	CN Loss	862185	q21.1	no	FRA13B
tr_13_set_2_clone_1	15	81501020	81588014	CN Loss	86995	q25.1	no	FRA2B/FRA22A
tr_17_set_2_clone_5	18	0	3152290	CN Loss	3152291	p11.32 - p11.31	no	
tr_18_set_2_clone_6	22	50864668	51304566	CN Gain	439899	q13.33	no	
tr_22_set_2_clone_10	5	26210976	52214687	CN Gain	26003712	p14.1 - q11.2	no	FRA5A/FRA5E
tr_5_set_1_clone_5	8	142486178	144510494	CN Loss	2024317	q24.3	no	FRA8D
tr_5_set_1_clone_5	8	146081828	146364022	CN Loss	282195	q24.3	no	FRA8D
tr_5_set_1_clone_5	22	16253076	51304566	CN Gain	35051491	q11.1 - q13.33	no	FRA2A/FRA2B/FRA22A/FRA22B
tr_6_set_1_clone_6	4	61577821	61784603	CN Loss	206783	q13.1	no	
tr_6_set_1_clone_6	7	155026587	155142658	CN Loss	116072	q36.2 - q36.3	no	FRA7I
tr_7_set_1_clone_7	2	172017814	187925525	CN Gain	15907712	q31.1 - q32.1	no	FRA2G/FRA2H
tr_8_set_1_clone_8	3	80776725	86007923	CN Loss	5231199	p12.2 - p12.1	no	